

TOTAL SYNTHESIS OF 5(S),12(S)- and 5(S),12(R)-DIHYDROXYEICOSA-
6(Z),8(E),14(Z)-TRIENOIC ACIDS, METABOLITES OF LEUKOTRIENE B₄

*Pendri Yadagiri, Sun Lumin, J.R. Falck**

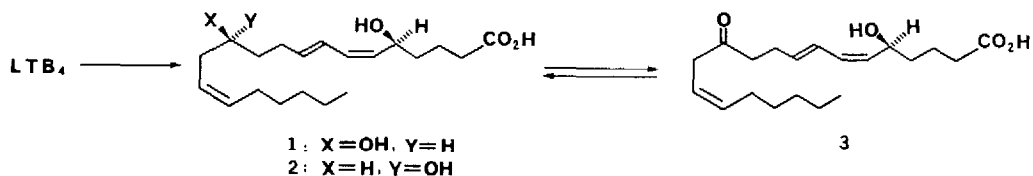
Departments of Molecular Genetics and Pharmacology, University of Texas
Southwestern Medical Center, Dallas, TX 75235 USA

*Armando Karara, and Jorge Capdevila**

Division of Nephrology, Vanderbilt University Medical Center,
Nashville, TN 37232 USA

Summary: The recently identified dihydro-leukotriene B₄ metabolite **1** and its C(12)-epi analogue **2** were prepared by Wittig coupling of segments derived from 2-deoxy-D-ribose and L-glutamic acid.

Leukotriene B₄ (LTB₄), a 5-lipoxygenase metabolite of arachidonic acid, is a potent endogenous mediator of macrophage and neutrophil activity and, consequently, may play a major role in inflammation and acute hypersensitivity.¹ Human neutrophils rapidly metabolize LTB₄ to biologically less active ω-oxidized products.² In contrast, other cell types³ primarily convert LTB₄ to a dihydro derivative recently assigned structure **1**.^{4a} Stable isotope studies suggest that the reductase acts directly on LTB₄ without activation of the triene system via a keto intermediate.⁴ It is also possible that the 12(R)-stereoisomer **2** is present since dihydro-LTB₄ (**1**) can be further metabolized⁴ in a reversible reaction to 12-oxo-dihydro-LTB₄ (**3**).

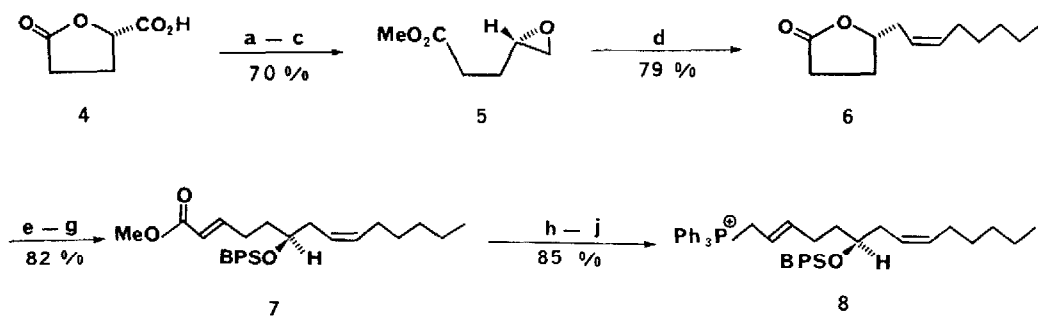


It is not known whether the reductase pathway plays a regulatory role⁵ leading to biologically inactive catabolites or is the source of a new family of eicosanoids with unique physiological properties. Support for the latter view is provided by the recent

characterization⁶ of dihydro-12-HETE in bovine cornea where it displays potent pro-inflammatory activity. To help address this and other urgent questions concerning dihydro metabolites as well as to clarify structure assignments, we report herein the asymmetric total synthesis of 1 and 2 using commercially available, chiral precursors.

Our synthesis of the fragment corresponding to C(7)-C(20) (Scheme I) commenced with borane-methyl sulfide reduction of carboxylic acid 4, readily accessible from L-glutamic acid⁷, followed by methanolysis of the derived primary tosylate to give epoxy-ester 5⁸ (70%). Addition of the higher order cuprate⁹ (1.1 equiv) generated from (Z)-1-iodo-1-heptene⁷ (9) to a 0.1 M solution of 5 with concomitant lactonization furnished 6 (79%), $[\alpha]_D^{23} + 17.7^\circ$ (c 3.7, MeOH); lit.¹⁰ $[\alpha]_D^{24} + 16.5^\circ$ (c 2.6, MeOH). Diisobutylaluminum hydride (DIBAL-H) reduction of 6 and trans-specific homologation of the resultant lactol⁶ with methyl (triphenylphosphoranylidene)acetate yielded ester 7¹¹ (82%) after silylation [TLC of 7: SiO₂, Et₂O/hexanes (1:6), R_f ~ 0.41]. The allylic alcohol obtained from 7 upon hydride reduction was transformed by standard procedures to Wittig salt 8¹² (85%) [TLC of 8: SiO₂, MeOH/CH₂Cl₂ (1:9), R_f ~ 0.29].

SCHEME I



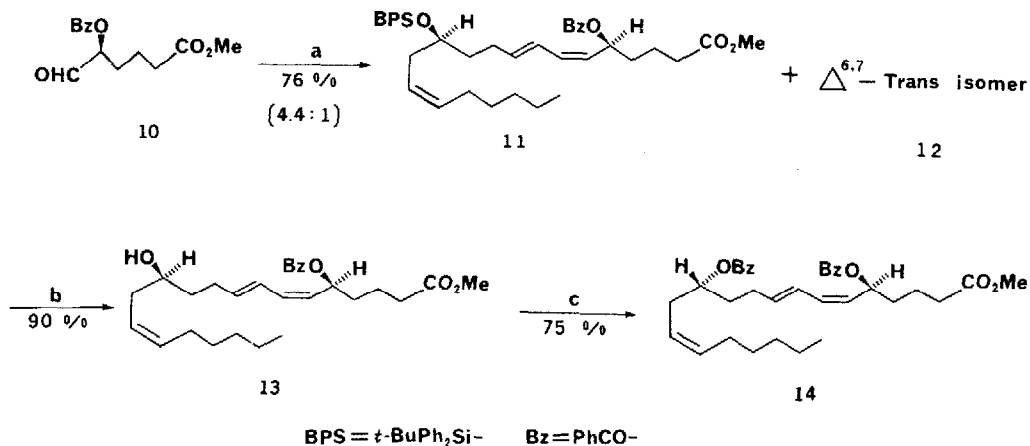
BPS = *t*-BuPh₂Si

^aBH₃·Me₂S, THF, 0° to 23°C over 2h. ^bTsCl, C₅H₅N/CH₂Cl₂ (1:2.5), 24°C, 6h. ^cNaOMe, MeOH, 20 min; AcOH to pH 7. ^d9, *n*-BuLi, Et₂O, -60°C, 20 min; CuCN, -78° to -45°C, 30 min; 5, -50°C, 10 min. ^eDIBAL-H, PhCH₃, -78°C, 3h. ^fPh₃PCHCO₂Me (1.5 equiv), PhCH₃, 23°C, 15h. ^g*t*-BuPh₂SiCl, Et₃N, DMAP, CH₂Cl₂, 45°C, 26h. ^hDIBAL-H, PhCH₃, -78°C, 1h. ⁱCBr₄, Ph₃P, CH₂Cl₂, 1h. ^jPh₃P, CH₃CN, 6h.

Union of methyl 5(S)-benzyloxy-5-formylpentanoate¹³ (10) with the ylide of 8 (1.2 equiv) afforded 11 (62%) and its Δ^{6,7}-*trans* isomer 12 (14%) after chromatographic purification [HPLC:Altex Ultrasphere Si (4.6 x 250 mm, 5 μ), EtOAc/hexane (5:95), 1 ml/min flow rate, R_t ~ 7.32 and 9.51 min for 11 and 12, respectively]. Desilylation of

11 with fluoride ion gave rise to alcohol 13¹⁴ (90%), $[\alpha]_{\text{D}}^{23} +160^\circ$ (c 1.68, CHCl_3), which was smoothly inverted to the 12(R)-isomer 14 (75%), $[\alpha]_{\text{D}}^{23} +141.7^\circ$ (c 1.2, CHCl_3), by the Mitsunobu procedure.¹⁵

SCHEME II



^ag, n-BuLi, THF/HMPA (4:1), -78°C , 1h; add 10 -78° to -50°C over 1.5h. ^b n-Bu₄NF, THF, 12h. ^cPhCO₂H, Ph₃P, EtO₂CNNCO₂Et, PhCH₃/pentane (1:1), 0°C , 1.5h.

Esters 13 and 14 were converted to 1 and 2, respectively, by saponification (LiOH, THF/H₂O 3:1), adjustment to pH 4.5, and extractive isolation¹⁶. Initial chromatographic comparisons revealed that natural dihydro-LTB₄ was an approximate 2:1 mixture of 1 and 2; none of the $\Delta^{6,7}$ -trans isomers of 1 was detected [SP-HPLC: Altex ROSIL (4.6 x 300 mm, 5 μ), hexane/isopropanol/acetic acid (933/66/1), 2 ml/min flow rate, $R_t \sim 16.0$, 15.4, and 20.8 min for 1, 2, and $\Delta^{6,7}$ -trans 1, respectively].^{4a} Additional results from investigations into the occurrence and pharmacological profile of this novel class of eicosanoids will be reported elsewhere.

Acknowledgment: Supported financially by grants from the USPHS NIH (GM 31278, 33541), the Robert A. Welch Foundation (I-782), and NATO (RG 85/0026). Funds for the purchase of a mass spectrometer were provided by NIH GM 16488.

References and Notes

1. B. Samuelsson, S.E. Dahlen, J.A. Lindgren, C.A. Rouzer, and C.N. Serhan, *Science* **237**: 1171-1176 (1987). P. Needleman, J. Turk, B.A. Jakschik, A.R. Morrison, and J.B. Lefkowitz, *Ann. Rev. Biochem.* **55**: 69-102 (1986).
2. Review: S. Hammarstrom, L. Orning, and K. Bernstrom, *Molecular and Cellular Biochemistry* **69**: 7-16 (1985).
3. W.S. Powell, *Biochem. Biophys. Res. Commun.* **145**: 991-998 (1987); V. Kaefer, M. Martin, J. Fauler, K.-H. Marx, and K. Resch, *Biochim. Biophys. Acta* **922**: 337-344 (1987).
4. (a) W.S. Powell (McGill University), personal communication; (b) W.S. Powell, Taipei Conference on Prostaglandin and Leukotriene Research, Taipei, Taiwan, Republic of China, April 22-24, 1988: Abstract S122, p. 87.

5. For an evaluation of the biological properties of dihydro-LTB₄ vis-a-vis LTB₄, see, V. Kaever, B. Damerou, K. Wessel, and K. Resch, *FEBS Lett.* **231**: 385-388 (1988).
6. R.C. Murphy, J.R. Falck, S. Lumin, P. Yadagiri, J.A. Zirrolli, M. Balazy, J.L. Masferrer, N.G. Abraham, and M.L. Schwartzman, *J. Biol. Chem.* **263**: 17197-17202 (1988).
7. U. Ravid, R.M. Silverstein, and L.R. Smith, *Tetrahedron* **34**: 1449-1452 (1978).
8. P.-T. Ho and N. Davies, *Synthesis*: 462 (1983).
9. B.H. Lipshutz, J. Kozlowski, and R.S. Wilhelm, *J. Amer. Chem. Soc.* **104**: 2305-2307 (1982).
10. W.H. Pirkle and P.E. Adams, *J. Org. Chem.* **44**: 2169-2175 (1979).
11. Satisfactory spectral data were obtained for all new compounds using chromatographically homogeneous samples.
12. Cf., D. Pirillo, L. Portaluppi, M. DiGiacomo, O. Azzolina, D. Vercesi, and G. Traverso, *IL Farmaco* **42**: 603-610 (1987).
13. Prepared as the methyl ester from 2-deoxy-D-ribose according to Y. Guindon, D. Delorme, C.K. Lau, and R. Zamboni, *J. Org. Chem.* **53**: 267-275 (1988).
14. Spectral data for **13**: ¹H NMR (CDCl₃, 250 MHz) δ 0.90 (t, J = 7.0 Hz, 3H), 1.21-1.42 (m, 6H), 1.59 (apparent q, J ~ 7Hz, 2H), 1.68-1.80 (m, 4H), 2.06 (apparent q, J ~ 6.8Hz, 2H), 2.24 (apparent q, J ~ 6.8Hz, 4H), 2.36 (apparent q, J ~ 7.0Hz, 2H), 3.55-3.68 (m, 1H), 3.65 (s, 3H), 5.35 (dd, J_{5,6} = 9.0, J_{6,7} = 11.0 Hz, 1H), 5.34-5.47 (m, 1H), 5.51-5.64 (m, 1H), 5.81 (dt, J_{9,10} = 6.8, J_{8,9} = 14.5 Hz, 1H), 5.91 (dt, J_{4,5} = 5.5, J_{5,6} = 9.0 Hz, 1H), 6.12 (dd, J_{7,8} = J_{6,7} = 11.0 Hz, 1H), 6.53 (dd, J_{7,8} = 11.0, J_{8,9} = 14.5 Hz, 1H), 7.42 (apparent t, J ~ 7.0Hz, 2H), 7.54 (apparent t, J ~ 7.0Hz, 1H), 8.03 (dd, J ~ 1 and 8Hz, 2H). Δ^{6,7}-trans isomer of **13**: δ 0.88 (t, J = 7.0 Hz, 3H), 1.20-1.40 (m, 6H), 1.50-1.63 (m, 2H), 1.69-1.86 (m, 4H), 2.05 (apparent q, J ~ 6.5Hz, 2H), 2.10-2.32 (m, 4H), 2.37 (apparent t, J ~ 7.0Hz, 2H), 3.53-3.68 (m, 1H), 3.68 (s, 3H), 5.30-5.44 (m, 2H), 5.47-5.61 (m, 3H), 5.60 (dd, J_{5,6} = 7.2, J_{6,7} = 15.0 Hz, 1H), 5.74 (dt, J_{9,10} = 6.8, J_{8,9} = 15.4 Hz, 1H), 6.05 (dd, J_{7,8} = 10.2, J_{8,9} = 15.4 Hz, 1H), 6.30 (dd, J_{7,8} = 10.2, J_{6,7} = 15.0 Hz, 1H), 7.43 (apparent t, J ~ 7.0Hz, 2H), 7.55 (apparent t, J ~ 7.0Hz, 1H), 8.04 (dd, J ~ 1 and 8Hz, 2H); [α]_D²³ + 45.6° (c 1.86, CHCl₃). **14**: δ 0.86 (t, J = 7.0 Hz, 3H), 1.19-1.36 (m, 6H), 1.64-1.91 (m, 6H), 1.97-2.12 (m, 2H), 2.18-2.32 (m, 2H), 2.36 (apparent t, J ~ 7.2Hz, 2H), 2.46 (apparent q, J ~ 6.8Hz, 2H), 3.65 (s, 3H), 5.12-5.22 (m, 1H), 5.36-5.56 (m, 4H), 5.81 (dt, J_{9,10} = 7.0, J_{8,9} = 14.5 Hz, 1H), 5.88 (dt, J_{4,5} = 5.5, J_{5,6} = 9.0 Hz, 1H), 6.10 (dd, J_{7,8} = J_{6,7} = 11.0 Hz, 1H), 6.50 (dd, J_{7,8} = 11.0, J_{8,9} = 14.5 Hz, 1H), 7.36-7.48 (m, 2H), 7.50-7.60 (m, 1H), 7.98-8.08 (m, 2H).
15. O. Mitsunobu, *Synthesis*: 1-28 (1981).
16. Mass spectrum of **1** as its TMS ether, methyl ester (PICl, CH₄): m/e 203, 229, 267, 285, 295, 317 (base), 345, 385, 407, 481, 495, 497 (M+1), 525 (M+29), 537 (M+41); MS (EI, 70 ev): m/e 147, 159, 203, 213, 229 (base), 265, 295, 305, 316, 338, 385, 395, 406, 481. The MS of **2** was virtually identical to that of **1**.

(Received in USA 30 September 1988)